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# New 17-Methyl-13-Octadecenoic and 3,16-Docosadienoic Acids from the Sponge *Polymastia penicillus*

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**Abstract** The phospholipid fatty acid composition of the North-East Atlantic sponge *Polymastia penicillus* (South Brittany, France) was investigated. Sixty fatty acids (FA) were identified as methyl esters (FAME) and *N*-acyl pyrrolidides (NAP) by gas chromatography–mass spectrometry (GC/MS), including eight  $\Delta 5,9$  unsaturated FA and three long-chain 2-hydroxylated FA. The major phospholipid FA were palmitic (14.3% of the total FA mixture), vaccenic (12.7%), 15(*Z*)-docosenoic (13.4%) and 5(*Z*),9(*Z*)-hexacosadienoic (13.3%) acids. In addition to the *iso*- and *anteiso*-branched saturated FA, several unusual short-chain branched saturated FA were identified. In addition to the known  $\Delta 5,9$  FA, and interestingly regarding their identification by GC–MS as *N*-acyl pyrrolidides, was the co-occurrence of unusual FA possessing a  $\Delta 3$ ,  $\Delta 4$  and  $\Delta 5$  double bond such as *iso*-4-pentadecenoic, *iso*-5-heptadecenoic, *anteiso*-5-heptadecenoic and two new compounds, not hitherto found in nature, namely 17-methyl-13-octadecenoic (0.8%) and 3,16-docosadienoic (1.1%) acids.

**Keywords** 3,16-Docosadienoic acid · Fatty acids · Phospholipids · *Polymastia* · Sponge ·

17-Methyl-13-octadecenoic · *N*-Acyl pyrrolidides · Non-methylene-interrupted fatty acids

## Abbreviations

amu	Atomic mass unit
FA	Fatty acid(s)
FAME	Fatty acid methyl ester(s)
GC/MS	Gas chromatography–mass spectrometry
MS	Mass spectrum (spectra)
NAP	<i>N</i> -Acyl pyrrolidide(s)
NMI	Non-methylene-interrupted
TLC	Thin layer chromatography

## Introduction

Marine sponges are primitive invertebrates with a remarkable ability to adapt to environmental changes. Thus, they have given rise to intensive investigations into the basic lipid components of their cell membranes since the pioneer studies [1, 2]. The phospholipid fatty acid composition is important for the functions of the cell membranes [3, 4]. In addition, sponge phospholipid classes have also been examined [3–5]. Moreover, sponges have proved to be a rich source of many unusual sterols and fatty acids (FA). Common polyunsaturated FA from the n-6 and n-3 series, such as eicosapentanoic acid EPA and docosahexaenoic acid DHA, are quite rare in sponge phospholipids, that instead usually contain very long-chain ( $C_{22}$ – $C_{34}$ ) non-methylene-interrupted (NMI) FA including the particular unsaturation pattern  $\Delta 5,9$  [3, 4, 6–11].

Sponges from the genus *Polymastia* are known to contain secondary metabolites with biological activity but there have been, in comparison, few investigations of

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their primary metabolites such as FA and sterols to date. Thus, organic extracts of *Polymastia janeirensis* demonstrated in vitro cytotoxicity against HT29 colorectal tumor cell lines [12]. *Polymastia croceus* has been farmed in order to produce quantities of bioactive compounds of medicinal interest [13, 14]. *Polymastia penicillus* is usually found in the North Atlantic in shallow waters and to depths of 600 m [15, 16]. The phospholipids of *P. gleneni*, collected around the Glenan Islands (South Brittany, France) were shown to contain an unique series of saturated 2-acetoxy long chain FA ranging from C<sub>22</sub> to C<sub>30</sub> chains [17].

As part of our ongoing investigations of sponge lipids [11, 18, 19], we present here the phospholipid FA composition of *P. penicillus* and the identification of several new and unusual  $\Delta$ 3,  $\Delta$ 4,  $\Delta$ 5 and  $\Delta$ 13 unsaturated FA.

## Materials and Methods

### Sponge Material

Samples of *Polymastia penicillus* were collected by hand (scuba) at a depth of 17 m, in the Etel River, Gulf of Morbihan, Brittany, France, in June 2005. Sponge identification was determined according to the recent classification [20, 21], and voucher specimens were deposited in the Marine Station of Endoume, Marseille, France. Like all Polymastiidae, this sponge is encrusting, hemispherical and cushion shaped, always with numerous papillae, but is characterized by the absence of a collagenous layer and by the absence of spicules in internal laminae, and spicules are tylostyles [15, 16].

### Extraction, Isolation and Analysis of Phospholipids

The collected specimens were immediately transported to the laboratory for immediate lipid extraction. The sponges were washed in seawater, carefully cleaned, cut in small pieces and steeped twice in dichloromethane/methanol (1:1, v/v) for 24 h at room temperature. The combined extracts yielded the crude total lipids. Total lipid content (wt.%) was determined as the ratio total lipids versus sum of sponge dried residue plus total lipids. Total lipids were rapidly subjected to lipid class separation. Phospholipids were separated from other lipids by column chromatography on silica gel (70–230 mesh) successively eluted with dichloromethane (neutral lipids), acetone (glycolipids) and methanol (phospholipids) [10, 11]. The phospholipid classes were investigated by thin-layer chromatography on silica gel 60F<sub>254</sub> plates (0.2 mm thickness, SDS Carlo Erba, Val de Reuil, France) and dichloromethane/methanol

(70:30, v/v) was the eluent with phospholipid standards purchased from Sigma (Saint-Quentin Fallavier, France).

### Preparation of Fatty Acid Methyl Esters and *N*-Acyl Pyrrolidides

Fatty acid methyl esters (FAME) were obtained by phospholipid transmethylation (40 min under reflux with 5% methanolic hydrogen chloride). *N*-Acyl pyrrolidide derivatives (NAP) were prepared by direct treatment of the FAME with pyrrolidine/acetic acid (10:1, v/v) for 2 h under reflux and purified by TLC on 0.5 mm silica gel layers using hexane/diethyl ether (1:2, v/v) [10, 22–26].

### Fourier Transform Infrared Spectroscopy

Infrared spectra were recorded by means of a Bruker Vector-22 Fourier transform infrared spectrometer (Bruker Spectrospin, Karlsruhe, Germany) on liquid films between NaCl plates. Spectra were collected over the wavenumber range 4,000 to 600 cm<sup>-1</sup>.

### Gas Chromatography–Mass Spectrometry Analyses

Gas chromatography–mass spectrometry (GC/MS) coupling experiments were performed on a Hewlett-Packard HP-5890 chromatograph linked to a HP-5989-A mass spectrometer (70 eV) equipped with a HP-9000-345 integrator, and using a 30 m × 0.32 mm i.d. fused silica capillary column coated with HP-1 (0.25 μm phase thickness). The carrier gas was helium. The temperatures of injector and source were held at 240 °C, and the temperature of the GC–MS interface was held at 290 °C.

The column temperature was programmed for FAME and NAP from 180 to 310 °C at 3 °C/min. Equivalent chain length (ECL) were determined for FAME and for NAP in some cases since the observed ECL values have proved to be very similar for both derivatives except for highly branched FA [25].

Mass spectral data for the novel fatty acid structures are presented below.

*17-Methyl-13-octadecenoic acid methyl ester* MS *m/z* (rel. int.) [M<sup>+</sup>] 310 (6.5), 279 (13.7), 227 (2.3), 211 (8.7), 194 (17.4), 185 (6.5), 171 (19.5), 152 (2.5), 139 (28.3), 125 (21.7), 111 (13.0), 97 (32.6), 95 (21.2), 87 (8.7), 83 (50.0), 81 (20.6), 74 (16.3), 69 (100), 55 (76.1), 43 (23.7), 41 (30.4).

*17-Methyl-13-octadecenoic acid pyrrolidide* MS *m/z* (rel. int.) [M<sup>+</sup>] 349 (16.1), 334 (4.5), 320 (1), 306 (5.4), 292 (4.5), 278 (3.4), 264 (4.5), 252 (2.7), 238 (2.7), 224 (4.5), 210 (3.6), 196 (3.4), 182 (3.6), 168 (4.5), 154 (1.8), 140

(3.6), 126 (23.8), 113 (100), 98 (11.6), 85 (4.5), 71 (9.8), 55 (9.8).

*3,16-Docosadienoic acid methyl ester* MS *m/z* (rel. int.) [M<sup>+</sup>] 350 (4.8), 337 (1.0), 319 (14.8), 304 (3.7), 290 (3.7), 276 (12.1), 261 (5.1), 247 (5.5), 191 (3.7), 171 (2.8), 163 (7.4), 150 (14.8), 135 (20.4), 123 (20.4), 115 (19.7), 109 (31.5), 95 (53.7), 87 (11.1), 81 (63.0), 71 (40.7), 69 (46.3), 67 (48.4), 55 (100), 43 (37.0), 41 (64.8).

*3,16-Docosadienoic acid pyrrolidide* MS *m/z* (rel. int.) [M<sup>+</sup>] 389 (32.2), 374 (2.5), 360 (4.1), 346 (4.2), 332 (11), 318 (19.5), 304 (7.6), 292 (5.1), 278 (3.4), 264 (7), 250 (6.8), 236 (6.8), 222 (10.2), 208 (11.9), 194 (8.5), 180 (4.2), 166 (8.5), 152 (61.9), 139 (13.6), 126 (30.5), 124 (9.3), 113 (100), 98 (30.5), 81 (8.5), 72 (30.5), 55 (8.5).

## Results and Discussion

Total lipids accounted for 5.3% of the dried sponge residues after extraction. Total phospholipids of *P. penicillus* accounted for 29.6% of the total lipids. As determined by thin layer chromatography, the most abundant phospholipids were phosphatidylserine, phosphatidylcholine and phosphatidylinositol, while phosphatidylethanolamine occurred at a significant level and phosphatidylglycerol was present at a lower amount. Acid methanolysis provided a complex phospholipid FA mixture. Sixty fatty acids (FA) were identified by gas chromatography–mass spectrometry (GC/MS) of their methyl esters (FAME) and *N*-acyl pyrrolidides (NAP) as shown in Table 1.

Many of the FA reported in Table 1 were identified by comparing their equivalent chain lengths (ECL) and their mass spectra (MS) with those of known standards. Unsaturated FA, mainly monoenes and dienes, accounted for almost 61% of the FA mixture. The Fourier transform infrared spectrum of the whole phospholipid FAME showed absorption at 3,004 cm<sup>-1</sup> (=C–H stretching), strong peaks at 2,923 and 2,852 cm<sup>-1</sup> (C–H stretching), 1,743 cm<sup>-1</sup> (C=O stretching, ester), significant peaks at 1,465 cm<sup>-1</sup> (CH<sub>2</sub> scissors deformation) and 720 cm<sup>-1</sup> (CH<sub>2</sub> rocking/C=C *cis*), and displayed no absorption in the region 940–980 cm<sup>-1</sup> (C=C–H out-of-plane), indicating the absence of (*E*)-unsaturation [27]. Thus, double bonds in major unsaturated phospholipid FA from *P. penicillus*, such as those at more than 1% have the (*Z*) configuration. It should be noted that the Δ<sup>5,9</sup> FA have been generally reported to have the (*Z*) configuration [3, 4].

Five long-chain dimethylacetal derivatives occurred as minor compounds in the phospholipid FA mixture. All their MS displayed the fragmentation ion *m/z* 75 ([CH<sub>3</sub>O)<sub>2</sub>–CH]<sup>+</sup> as the base peak, and a appreciable [M–MeO]<sup>+</sup> ion [26, 28]. Unfortunately, complete identification

**Table 1** Phospholipid fatty acids identified from the sponge *Poly-mastia penicillus*

Fatty acid (symbol)	ECL <sup>1</sup> (HP-1)	Abundance (wt%)
Saturated fatty acids (SFA)		
Dodecanoic (12:0)	12.00	0.6
Tetradecanoic (14:0)	14.00	1.3
10-Methyltetradecanoic (br-15:0)	14.58	2.7
13-Methyltetradecanoic ( <i>i</i> -15:0)	14.60	0.4
Pentadecanoic (15:0)	15.00	0.3
12-Methylpentadecanoic (br-16:0)	15.52	2.1
14-Methylpentadecanoic ( <i>i</i> -16:0)	15.62	0.6
Hexadecanoic (16:0)	16.00	4.3
15-Methylhexadecanoic ( <i>i</i> -17:0)	16.60	1.2
14-Methylhexadecanoic ( <i>ai</i> -17:0)	16.71	1.8
Heptadecanoic (17:0)	17.00	0.2
Octadecanoic (18:0)	18.00	2.6
15-Methylnonadecanoic (br-20:0)	19.60	0.6
18-Methylnonadecanoic ( <i>i</i> -20:0)	19.66	0.5
17-Methylnonadecanoic ( <i>ai</i> -20:0)	19.72	0.3
Eicosanoic (20:0)	20.00	1.0
19-Methyleicosanoic ( <i>i</i> -21:0)	20.63	0.6
Tricosanoic (23:0)	23.00	0.5
Hexacosanoic (26:0)	26.00	0.6
Total SFA: 32.2%		
2-Hydroxy fatty acids (HFA)		
2-Hydroxytetracosanoic (2-OH-24:0)	25.23	3.4
2-Hydroxypentacosanoic (2-OH-25:0)	26.22	0.8
2-Hydroxyhexacosanoic (2-OH-26:0)	27.14	2.8
Total HFA: 7.0%		
Monounsaturated fatty acids (MFA)		
13-Methyl-4-tetradecenoic ( <i>i</i> -15:1)	14.28	0.4
( <i>Z</i> )-9-Hexadecenoic (16:1)	15.65	1.6
15-Methyl-5-hexadecenoic ( <i>i</i> -17:1)	16.28	0.7
14-Methyl-5-hexadecenoic ( <i>ai</i> -17:1)	16.40	0.4
( <i>Z</i> )-9-Heptadecenoic (17:1)	16.81	1.0
5-Octadecenoic (18:1)	17.49	0.6
( <i>Z</i> )-11-Octadecenoic (18:1)	17.82	12.7
17-Methyl-13-octadecenoic ( <i>i</i> -19:1) <sup>2</sup>	18.21	0.8
9-Nonadecenoic (19:1)	18.70	0.8
12-Nonadecenoic (19:1)	18.82	0.6
11-Eicosenoic (20:1)	19.76	0.5
13-Eicosenoic (20:1)	19.82	0.4
x <sub>1</sub> -Heneicosenoic (21:1)	20.80	0.6
x <sub>2</sub> -Heneicosenoic (21:1)	20.84	0.4
( <i>Z</i> )-15-Docosenoic (22:1)	21.87	13.4
16-Tetracosenoic (23:1)	22.82	0.3
( <i>Z</i> )-15-Tetracosenoic (24:1)	23.62	1.7
( <i>Z</i> )-17-Tetracosenoic (24:1)	23.81	1.5
19-Tetracosenoic (24:1)	23.88	0.7
( <i>Z</i> )-17-Hexacosenoic (26:1)	25.61	2.3

**Table 1** continued

Fatty acid (symbol)	ECL <sup>1</sup> (HP-1)	Abundance (wt%)
Total MFA: 41.4%		
Diunsaturated fatty acids (DFA)		
(5Z,9Z)-5,9-Octadecadienoic (18:2)	17.40	1.4
(9Z,12Z)-9,12-Octadecadienoic (18:2)	17.62	0.8
(5Z,9Z)-5,9-Eicosadienoic (20:2)	19.39	0.3
11,15-Eicosadienoic (20:2)	19.60	0.4
3,16-Docosadienoic (22:2) <sup>2</sup>	21.55	1.1
(5Z,9Z)-5,9-Tetracosadienoic (24:2)	23.44	0.9
(5Z,9Z)-5,9-Pentacosadienoic (25:2)	24.50	0.7
(5Z,9Z)-5,9-Hexacosadienoic (26:2)	25.52	13.3
Total DFA: 18.9%		

Identified minor (<0.2%) fatty acids: *i*-14:0 (13.64), 10-Me-16:0 (16.52), 13-Me-16:0 (16.56), 14-Me-17:0 (17.55), Phytanic (17.74), 15-20:1 (19.88), 23:0 (23.00), 5,9,*x*-25:3 (24.20), br-5,9-25:2 (24.22), 5,9-27:2 (26.48)

<sup>1</sup> FAME

<sup>2</sup> Not previously found in nature

of these derivatives failed due to their very small amounts and that they were co-eluted with other compounds. It is known that they are obtained from particular phospholipids named plasmalogens (1-*O*-alk-1'-enyl-2-acyl-glycerophospholipids) of great biological interest, and were found in some sponges [29, 30]. Dimethylacetals are produced from fatty aldehydes, which are liberated from plasmalogen vinyl ether bonds on methanolic acid hydrolysis.

### Saturated Fatty Acids

Twenty-five saturated FA of Table 1 were identified accounting for more than 32% of the total FA, including fourteen monobranched ones from C<sub>14</sub> to C<sub>21</sub> (almost 11%).

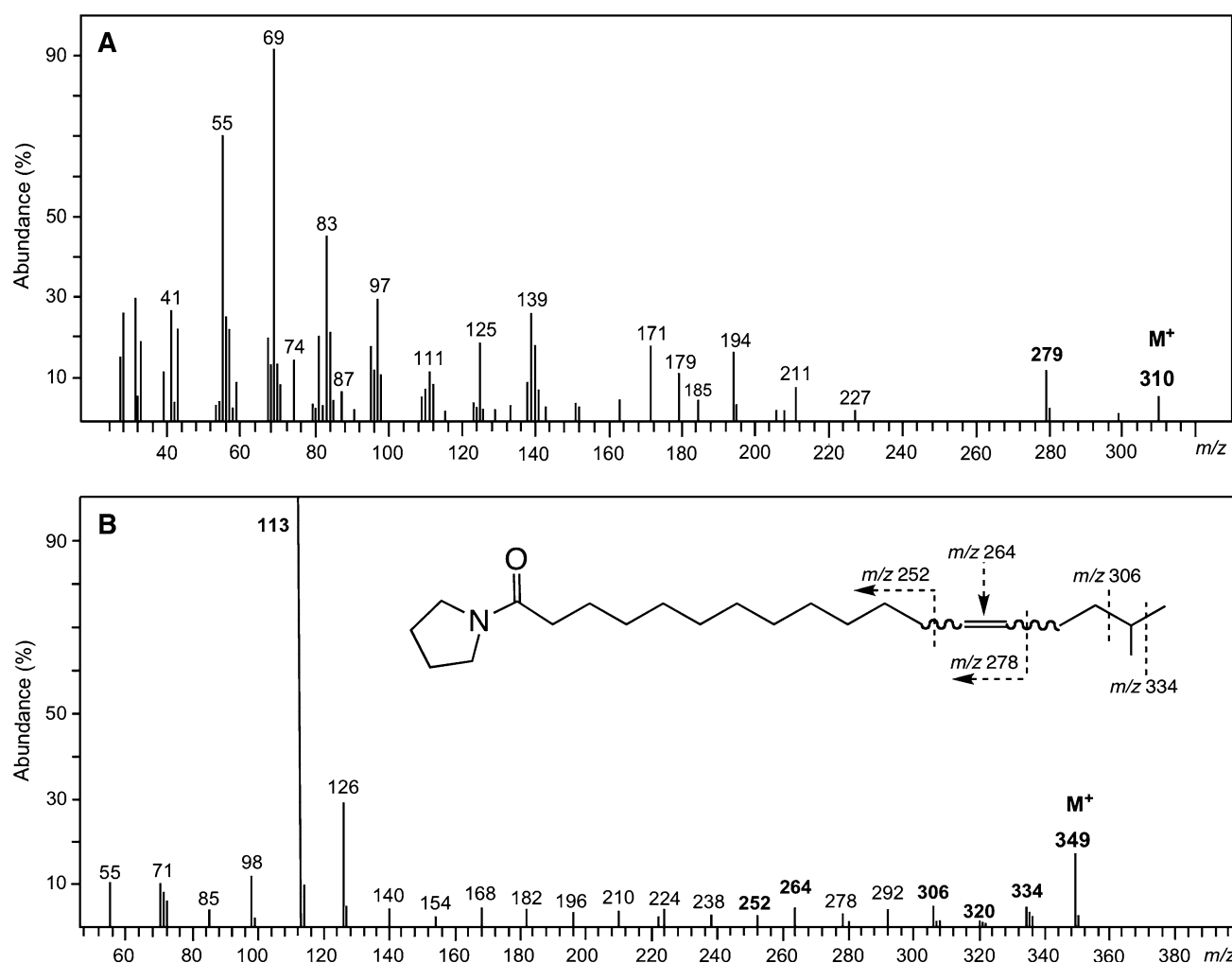
The highly branched phytanic acid present as trace was readily identified as methyl ester with an ECL value of 17.74 and diagnostic fragmentation ions including the molecular ion at *m/z* 326 and a base peak at *m/z* 101 (C-3 methyl branching), as previously described [25, 26]. It was the sole isoprenoid FA identified in the present mixture.

As usually, *iso*- and *anteiso*-branched FA were identified as FAME and NAP from their ECL values and their MS typical of saturated FA (*m/z* 74 base peak, McLafferty, *m/z* 87). The MS of their NAP displayed the corresponding molecular ion and a diminished peak for the ions [M-CH<sub>3</sub>-CH<sub>2</sub>]<sup>+</sup> and [M-CH<sub>3</sub>-2CH<sub>2</sub>]<sup>+</sup>, respectively [10, 18, 19, 22, 24].

In addition to the *iso*- and *anteiso*-branched FA, seven various monomethyl-branched FA occurred in phospholipids of *P. penicillus*. Six of them with short-chains were

detected as FAME from their ECL values of 14.58, 15.52, 16.52, 16.56, 17.55 and 19.60, respectively, and from their MS that exhibited molecular ions at *m/z* 256, 270, 284 (two compounds) and 326, respectively. Thus, we dealt with branched pentadecanoic, hexadecanoic, heptadecanoic, eicosanoic acid structures. The MS of the 15:0 pyrrolidide showed a very low fragmentation ion at *m/z* 224 (C<sub>10</sub> fragment) and enhanced peaks at *m/z* 210 and 238 indicating a methyl branch at C-10 [10, 22, 24]. Thus, the unusual 10-methyltetradecanoic acid was identified. Another branched 17:0 acid was identified as FAME and as NAP. As FAME its MS showed the molecular ion at *m/z* 284 and an expected ion at *m/z* 227 of very low intensity, while the ions at *m/z* 213 and 241 were highly enhanced (methyl branch at C-13) [24, 29]. The branched 17:0 acid was then identified (FAME) as 13-methylhexadecanoic acid and this structure was confirmed by the MS of the NAP which showed a molecular ion at *m/z* 323. The expected ion *m/z* 266 was lacking, while the ions at *m/z* 252 and *m/z* 280 were highly enhanced (methyl branching at C-13). Mass spectra of NAP of 13-methyl- and 15-methyl-branched saturated FA were shown in a recent report [29]. The quite unusual 12-methylpentadecanoic, 14-methylheptadecanoic and 15-methylnonadecanoic acids were characterized as methyl esters since their MS showed the molecular ions at *m/z* 270, 298 and 326 (16:0, 18:0 and 20:0 structures) [24, 29]. Among these FA, the branched 20:0 acid (FAME) was identified as 15-methylnonadecanoic acid since the expected ion at *m/z* 255 was lacking, while the ions at *m/z* 241 and 269 were present (methyl branch at C-14) [22, 29].

*Iso*- and *anteiso* FA are typical for the occurrence of bacteria and the other monomethyl-branched short-chain FA probably have the same origin [31–34], although one of the most frequent bacterial phospholipids, namely phosphatidylglycerol, accounted here for a relatively low percentage [3, 4]. Branched-chain FA (*iso* and *anteiso*) are characteristic for gram-positive bacteria [34]. The 13-methylhexadecanoic acid was described in previous reports [22, 29]. Branched saturated FA (C<sub>15</sub>–C<sub>19</sub>) have been used as biomarkers for bacteria including anaerobic bacteria and the sulfate-reducing bacteria [33–37]. The 10-methyl-14:0, 10-methyl-15:0 and 10-methyl-16:0 acids were found in bacteria [33–35], and it was assumed that methyl branching on the tenth carbon atom in the acyl chain seems specific for actinomycetes [33, 38]. The 10-methylhexadecanoic acid was observed in a *Cinachyrella* sponge [10] and in a pioneering report on the sponge *Aplysina fistularis* [4]. The occurrence of bacteria in sponges has already been evidenced and studied in several reports [39, 40].



**Fig. 1** Mass spectra of methyl ester (a) and pyrrolidide derivative (b) of 17-methyl-13 octadecenoic acid from *Polymastia penicillus*

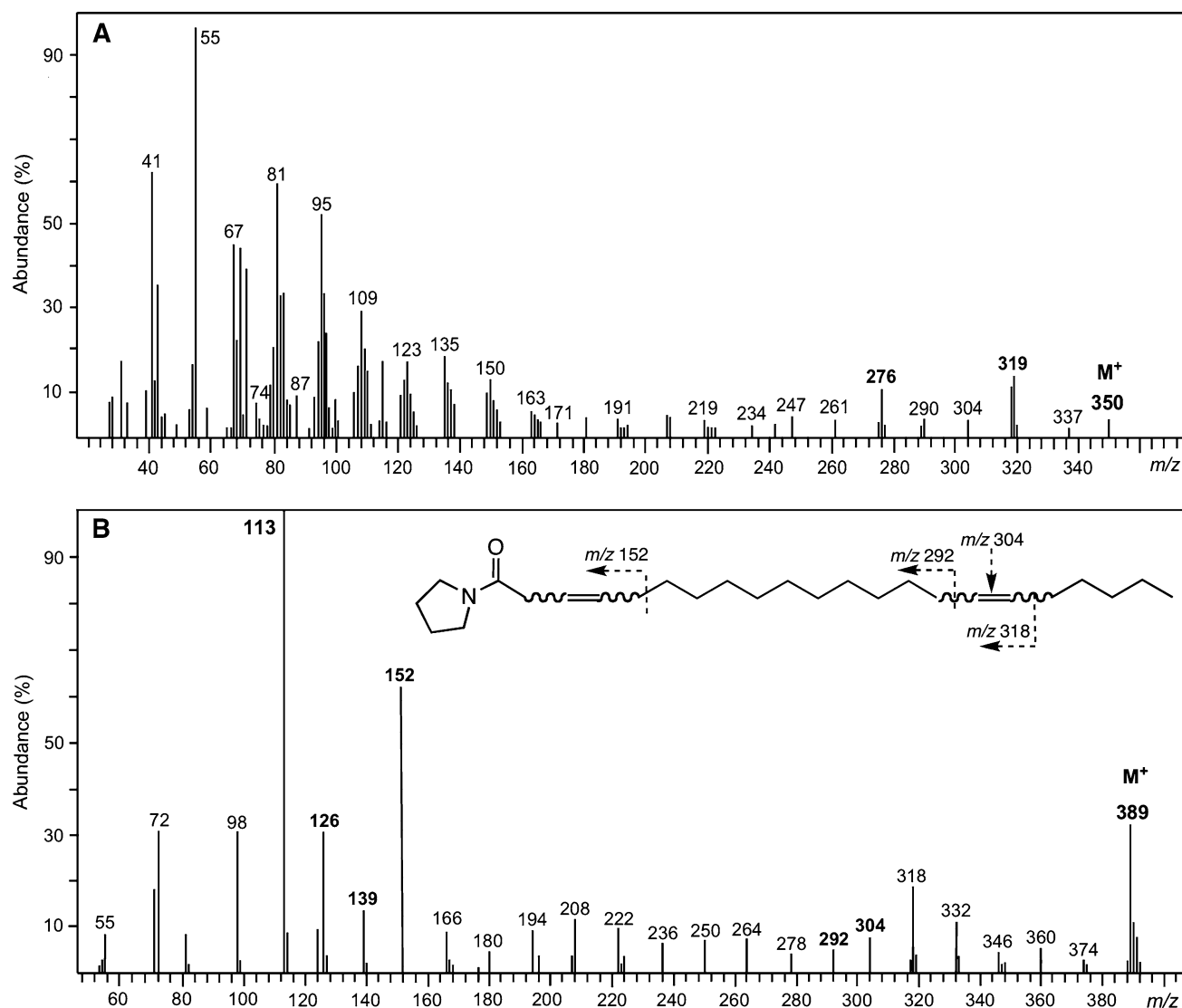
## 2-Hydroxylated Fatty Acids

Three 2-hydroxylated FA (7.0% of FA mixture; C<sub>24</sub>, C<sub>25</sub> and C<sub>26</sub>) were readily identified by GC/MS as FAME since their MS displayed diagnostic fragmentation ions at *m/z* 90 (McLafferty) and 103, and the corresponding molecular ions at *m/z* 398, 412 and 426, respectively. Diagnostic ions of 2-hydroxy acids have been described in previous reports such as those on series of 2-hydroxylated FA found in two sponges of the family Suberitidae [29] and in two Caribbean sponges [31]. The M<sup>+</sup> and *m/z* 90 and 103 ions were accompanied by the corresponding related ions [M–MeOH]<sup>+</sup> at *m/z* 366, *m/z* 380 and *m/z* 394, respectively, and [M–COOMe]<sup>+</sup> at *m/z* 339, *m/z* 353 and *m/z* 367, respectively. Mass spectra of NAP derivatives showed the corresponding molecular ions and the major ions at *m/z* 129 (McLafferty, base peak), *m/z* 113, *m/z* 100 and *m/z* 98, thus confirming the hydroxylated structures.

## Monounsaturated Fatty Acids

Twenty-one monounsaturated FA listed in Table 1 were identified, accounting for more than 41% of the total FA, including four *iso*- and *anteiso*-branched FA. Three unusual short-chain monoenoic acids were readily identified in phospholipids of *P. penicillus*.

A short-chain acid was readily identified as a branched pentadecenoic acid (0.4%; ECL 14.28) since its NAP displayed a molecular ion at *m/z* 293 (15:1), and major fragment ions at *m/z* 126 and *m/z* 152, a peak at *m/z* 138 of low intensity, a very intense peak at *m/z* 166 (almost 70% of the base peak *m/z* 113; Δ4), and lacking the expected *m/z* 264, the ions at *m/z* 250 and *m/z* 278 being enhanced. Thus, we identified the *iso*-4-pentadecenoic acid, which was early observed in phospholipids from *Suberites massa* [29]. In addition, some examples of MS of NAP of Δ4 FA are shown in Christie's homepage "[www.lipidlibrary.co.uk](http://www.lipidlibrary.co.uk)"



**Fig. 2** Mass spectra of methyl ester (a) and pyrrolidide derivative (b) of 3,16-docosadienoic acid from *Polymastia penicillus*

[23, 24, 26]. Another  $\Delta 4$  monounsaturated FA occurred in the freshwater sponge *Baicalospongia bacilifera*, namely *iso*-4-octadecenoic acid [41]. The latter sponge also contained  $\Delta 5$  FA as shown with the use of the iodolactonization method [41].

Two closely related FA were identified as monounsaturated and monomethyl-branched FA since their MS showed a molecular ion (as FAME) at  $m/z$  282, and that their ECL values were 16.28 and 16.40. Mass spectra of their NAP showed the expected molecular ion at  $m/z$  321 and a gap of 12 amu between an appreciable  $m/z$  140 and 152, while the ion at  $m/z$  166 was enhanced, indicative of  $\Delta 5$  unsaturation. The first br-17:1 acid was identified as 15-methylhexadecenoic (*iso*-17:1) acid because of the low relative intensity of the fragment ion at  $m/z$  294 ( $C_{15}$ ), while the ions at  $m/z$  278 ( $C_{14}$ ) and 306 ( $C_{16}$ ; M- $CH_3$ ) were enhanced. The second br-17:1 acid was identified as

14-methylhexadecenoic (*anteiso*-17:1) acid due to the low relative intensity of the fragment ion at  $m/z$  278 ( $C_{14}$ ), while the ions at  $m/z$  264 ( $C_{13}$ ) and 292 ( $C_{15}$ ) were enhanced. Thus, we characterized two not-so-common FA, namely 15-methyl-5-hexadecenoic and 14-methyl-5-hexadecenoic acids. These *iso* and *anteiso* monounsaturated  $\Delta 4$  or  $\Delta 5$  FA probably originated from associated bacteria. Present at low growth temperatures in bacteria from the genera *Bacillus*, *Brevibacter* and *Streptomyces*, the *iso*- $\Delta 5$ -hexadecenoic, *anteiso*- $\Delta 5$ -pentadecenoic and *anteiso*- $\Delta 5$ -heptadecenoic acids were identified as methyl esters [42].

Another branched monoenoic FA was detected by GC-MS as FAME and characterized as NAP (Fig. 1).

This acid (FAME) showed a molecular ion at  $m/z$  310, and a (M- $CH_3O$ ) $^+$  at  $m/z$  279, corresponding to a 19:1 acid structure, and an ECL value of 18.21, implying methyl branching (Fig. 1a). The usual peaks for FA methyl esters

at  $m/z$  74 and  $m/z$  87 were present as minor fragment ions. Other abundant fragments were not relevant for the characterization of structure. The molecular ion peak of the corresponding NAP was present at  $m/z$  349 confirming the 19:1 structure (Fig. 1b). The gap of 12 amu between homologous fragment ions at  $m/z$  252 ( $C_{12}$ ) and  $m/z$  264 ( $C_{13}$ ) indicated  $\Delta 13$  unsaturation. A diminished peak at  $m/z$  320 ( $C_{17}$ ) flanked by an elevated peak at  $m/z$  306 ( $C_{16}$ ), and an appreciable peak at  $m/z$  334 ( $C_{18}$ ), indicated a methyl branch at C-17. Thus, we identified the new 17-methyl-13-octadecenoic acid, which to our knowledge, has not been reported from any natural source.

### Diunsaturated Fatty Acids

Ten dienoic FA were identified accounting for about 19% and including the 9,12-octadecadienoic acid (0.8%), a series of eight FA possessing the 5,9-unsaturation pattern common in sponges, and two other interesting NMI FA.

The FA with the 5(*Z*),9(*Z*)-unsaturation, ranged from  $C_{18}$  to  $C_{27}$ , were readily characterized by GC-MS. Mass spectra of all their methyl esters displayed the corresponding molecular ion and the ion at  $m/z$  81 (base peak), while spectra of their NAP displayed a significant ion at  $m/z$  180 arising from the activated double allylic cleavage. Mass spectra of both derivatives showed corresponding molecular ions. They had been already found in sponges [4, 6–11, 18, 19, 26, 29, 31, 32]. It is now well known that lipids of most sponge species contain various  $\Delta 5$  olefinic FA mainly including  $\Delta 5,9$  FA. Interestingly,  $\Delta 5,9$  FA displayed antiprotozoal activity, antitumor activity, and topoisomerase I inhibitory activity [43, 44]. Only one triunsaturated FA occurred as a minor component in the FA mixture and was identified as 5,9,*x*-pentacosatrienoic acid since its methyl ester (ECL of 24.20) showed the molecular ion at  $m/z$  390 and the base peak at  $m/z$  81.

A first additional NMI FA (0.3%) had an ECL value of 19.60 as methyl ester and a molecular ion at  $m/z$  322 indicative of a 20:2 acid structure. The MS of the NAP showed a molecular ion at  $m/z$  361 (20:2 acid structure). The two double bonds were located by the 12 amu differences between the peaks at  $m/z$  224 ( $C_{10}$ ) and  $m/z$  236 ( $C_{11}$ ), indicating of  $\Delta 11$  unsaturation, and between the peaks at  $m/z$  278 ( $C_{14}$ ) and  $m/z$  290 ( $C_{15}$ ), indicating  $\Delta 15$  unsaturation. Thus, we dealt with the rare 11,15-eicosadienoic acid, which was firstly reported from the sponge *Amphimedon complanata* [45] and observed in *Stylissa carteri* [19]. This FA has been considered as a biosynthetic intermediate of the unusual dienoic FA 17,21-hexacosadienoic and 19,23-octacosadienoic acids [10].

The second interesting dienoic NMI FA (1.1%) as a methyl ester had an ECL value of 21.55 and a molecular ion at  $m/z$  350, and elevated ions at  $m/z$  319 ( $[M-74]^+$ ) and

$m/z$  276 ( $[M-31]^+$ ), indicative of a 22:2 acid structure (Fig. 2a).

The usual peaks for FA methyl esters at  $m/z$  74 and  $m/z$  87 were present as minor fragment ions. The relatively abundant ions at  $m/z$  304 ( $C_{16}$ ),  $m/z$  290 ( $C_{15}$ ), and  $m/z$  276 ( $C_{14}$ ) confirmed a 22:2 FA structure. The MS of the NAP showed a molecular ion at  $m/z$  389 (22:2 structure) (Fig. 2b). A first double bond was readily located by the 12 amu gap between the peaks at  $m/z$  292 ( $C_{15}$ ) and  $m/z$  304 ( $C_{16}$ ), indicating of  $\Delta 16$  unsaturation. The second double bond was located with the sequence of  $m/z$  113 (base peak),  $m/z$  126,  $m/z$  139,  $m/z$  152 (the second-most prominent ion at more than 60% of the relative intensity) and  $m/z$  166, indicating a  $\Delta 3$  unsaturation according to Andersson [23, 24]. It should be noted that the ion at  $m/z$  152 was the second-most prominent ion in the MS of the NAP of the novel 3(*Z*),9(*Z*),12(*Z*)-octadecatrienoic acid that also exhibited the two appreciable  $m/z$  126 and  $m/z$  139 [26, 46]. Also available on the Christie's homepage, MS of the NAP of the 3(*E*),9(*Z*)-octadecadienoic acid allows another useful comparison [26]. Indeed, the  $m/z$  152 ion is also the second-most prominent ion but there is a prominent ion at  $m/z$  124 suggesting a probable isomerization to 2,9-18:2 during the derivatization step. Thus, MS of the NAP of 2,6-18:2 shows intense ions at  $m/z$  124 without the  $m/z$  126, and at  $m/z$  139 (base peak) [26]. In addition, MS of 2-16:1 and 2-18:1 show similar features with a major ion at  $m/z$  152, a prominent ion at  $m/z$  124 and a minor ion at  $m/z$  126 [26].

In the MS of the 22:2 (NAP), from *P. penicillus*, the ion at  $m/z$  126 is intense in accordance with a  $\Delta 3$  double bond. Nevertheless, the presence of the ion at  $m/z$  124 with a lower intensity than the ion at  $m/z$  126 in the SM depicted in Fig. 2 suggested a possible partial isomerization  $\Delta 3$  to  $\Delta 2$  [26]. Thus, we identified the new 3,16-docosadienoic acid, which to our knowledge, has not been reported from any natural source.

In summary, phospholipids from the sponge *P. penicillus* contained a lot of monomethyl-branched monoenoic FA, several  $\Delta 5,9$  FA, and two novel FA, namely 17-methyl-13-octadecenoic and 3,16-docosadienoic.

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