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Synthesis of nitro-aryl functionalised 4-amino-1,8-naphthalimides and their evaluation as fluorescent hypoxia sensors

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Abstract:

Fluorescent sensors are a vital research tool, enabling the study of intricate cellular processes in a sensitive manner. The design and synthesis of responsive and targeted probes is necessary to allow such processes to be interrogated in the cellular environment. This remains a challenge, and requires methods for functionalisation of fluorophores with multiple appendages for sensing and targeting groups. Methods to synthesise more structurally complex derivatives of fluorophores will expand their potential scope. Most known 4-amino-1,8-naphthalimides are only functionalised at imide and 4-positions, and structural modifications at additional positions will increase the breadth of their utility as responsive sensors. Here we evaluate methods for the incorporation of a hypoxia sensing group to 4-amino-1,8-naphthalimide. We developed an intermediate that allowed us to incorporate a sensing group, targeting group, and ICT donor to the naphthalimide core in a modular fashion. Synthetic strategies for attaching the hypoxia sensing group and how they affected the fluorescence of the naphthalimide were evaluated by photophysical characterisation and time-dependent density functional theory. We then rationally designed an extracellular hypoxia probe that could selectively image the hypoxic and necrotic region of tumour spheroids. Our results demonstrate the versatility of the naphthalimide scaffold and expand its utility. This approach to probe design will enable the flexible, efficient generation of selective, targeted fluorescent sensors for various biological purposes.

Introduction:

Fluorescent sensing, particularly when applied to bioimaging, is a vital tool for studying chemical processes in biological systems. Many unanswered biochemical questions require selective,

targeted, responsive fluorescent sensors and their development remains challenging.^[1] Responsive fluorescent sensors typically comprise a sensing moiety tethered to a fluorophore, often through a conjugated linker. A particular challenge in the rapid development of new fluorescent sensors is the identification of fluorophore precursors to which sensing groups can be readily conjugated. To achieve this, it is important to identify fluorophore-linker scaffolds that enable facile conjugation of sensing groups, and for which the analyte induces a measurable fluorescence response.

4-Amino-1,8-naphthalimides are donor-acceptor molecules with an internal charge-transfer (ICT) excited state, and typically display strong fluorescence emission.^[2] They are commonly used in bioimaging due to their chemical stability, photostability, high quantum yields, and large Stokes shifts.^[3] Responsive 4-amino-1,8-naphthalimide based sensors have been reported for several analytes including metal ions,^[4] anions,^[5] and reactive small molecules.^[6] Functionalisation at the imide or the 4-position predominates the literature (Figure 1), with additional functionalisation uncommon. Because of the limited scope for derivatisation to date, naphthalimides remain an underexplored class of fluorophore, and in order to maximise the potential of naphthalimides as responsive fluorescent sensors, it is essential to develop methods for functionalisation at other positions. Importantly, we identified the need to determine robust synthetic methods for introducing a broad range of sensor groups. This will allow rapid access to more structurally complex derivatives and enable more efficient development of a set of fluorescent sensors. We were particularly interested in a third point of attachment which could be functionalised orthogonally to the imide and 4-position.

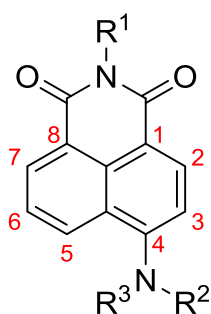


Figure 1: 4-Amino-1,8-naphthalimide structure

We have previously reported several substituted 4-amino-1,8-naphthalimide derivatives with various simple structural modifications and found that the addition of substituents, particularly at the 6-position, expanded the range of emission wavelengths available.^[7] An additional electron donating group at this position was shown to have a large impact as a secondary charge transfer (CT) donor, making it a promising position for incorporation of sensing groups. Among the compounds previously synthesised was a 6-bromo analogue, which had the potential to be used as a synthetic

handle. This work aims to assess the synthetic strategies that are most promising for attaching sensing groups at this position, both in terms of optimal photophysical properties, and synthetic considerations.

We envisaged that the incorporation of nitro-aryl substituents onto the 4-amino-1,8-naphthalimide scaffold would provide a suitable model system in which to examine the above properties, anticipating that the resulting molecules would be useful probes for sensing hypoxic environments, since aromatic nitro compounds are known to be reduced to the corresponding aniline in hypoxic conditions.^[8] Nitro groups generally quench fluorescence,^[9] whereas in amine-containing fluorophores, strong fluorescence is common. This difference in fluorescence properties, coupled with the ease of reduction of the nitro group makes this an attractive strategy for the design of fluorogenic hypoxia probes and an ideal model system for our studies.^[10]

Hypoxia develops in advanced solid tumours due to uncontrolled proliferation of cancer cells, resulting in regions that are distant from blood vessels, or possess inadequate tumoral vasculature, and therefore are starved of oxygen.^[11] The development of these hypoxic regions can aid in tumorigenicity and promote changes that lead to disease progression, with acute hypoxia being associated with more aggressive tumour phenotypes and resistance to treatment.^[12] Fluorescence imaging is a useful technique for imaging hypoxic regions of tumours as it is non-invasive, highly sensitive, and can provide spatiotemporal resolution. Other commonly-used methods for measuring hypoxia such as microelectrodes are invasive techniques.^[13] To investigate the ease with which the nitro-aryl hypoxia sensing groups could be introduced onto the 4-amino-1,8-naphthalimide scaffold, together with the impact of different linkers on the photophysical properties of these systems we prepared a small series of 6-substituted-4-amino-1,8-naphthalimides and report here our investigations of these as fluorescent sensors of hypoxic microenvironments.

Results/Discussion:

Design/Synthesis

We chose to incorporate the aryl-nitro functionality *via* four robust methodologies that are commonly used in synthetic chemistry, with readily available substrates: directly conjugated by a Suzuki-coupling, with an alkyne linker using a Sonogashira coupling, a triazole linker using Cu-catalysed azide-alkyne cycloaddition (CuAAC) click reaction, and by Buchwald-Hartwig amination. We designed four nitro-compounds (**1NO₂** - **4NO₂**), and their reduced aniline analogues (**1NH₂** - **4NH₂**) (Figure 2), to investigate how the aryl-nitro functionality is incorporated impacts the emission.

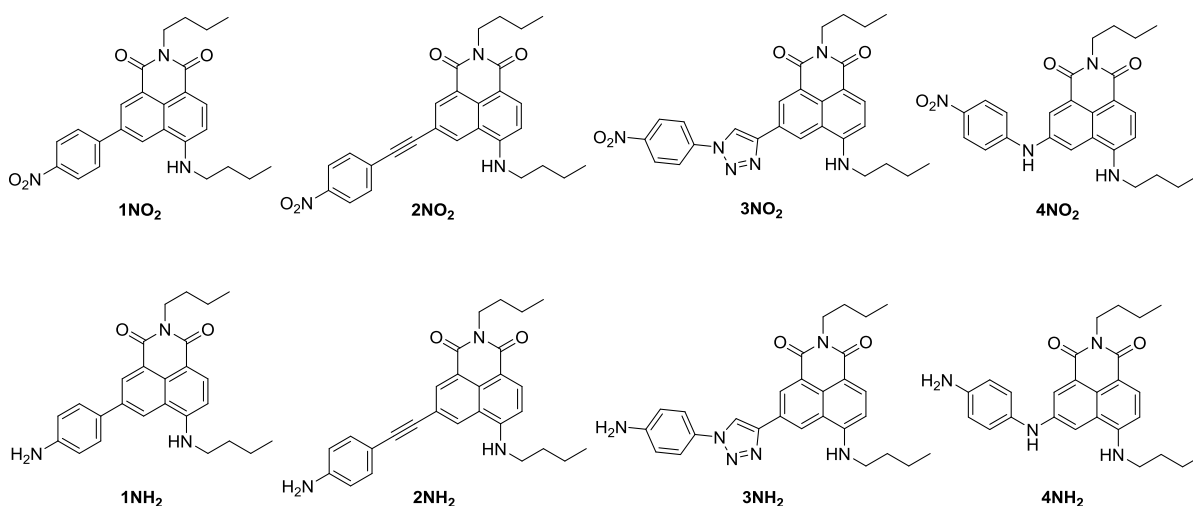
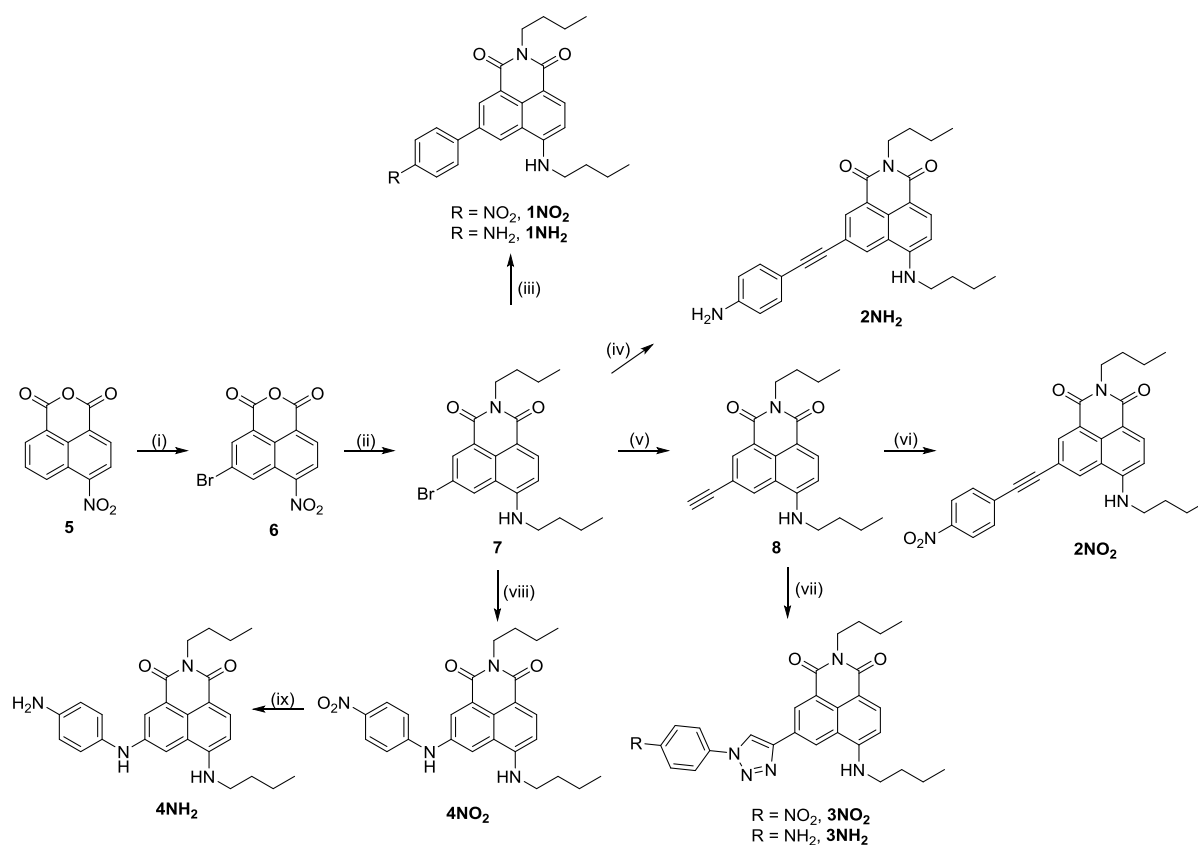


Figure 2: Structure of 6-aryl-nitro naphthalimides **1 – 4NO₂** and their reduced aniline derivatives **1 – 4NH₂**

The reported synthesis of 6-bromonaphthalimide **7** contains a non-selective nitration and is low yielding for the desired regioisomer.^[7] We therefore developed an alternative route to **7** with selective bromination of 4-nitro-1,8-naphthalic anhydride as the initial step. Commercially available 4-nitronaphthalic anhydride **5** was brominated regioselectively at the 6-position using NBS in strongly acidic conditions, giving the desired product **6** with no need for chromatographic purification. This substituted anhydride **6** is a key intermediate and allows for facile and selective functionalisation at each of the 4-, 6- and anhydride positions of the scaffold. Treatment with excess *n*-butylamine in EtOH gave bromo-naphthalimide **7**, in good overall yield leaving the 6-bromide handle for further functionalisation. Suzuki coupling of **7** with 4-nitrophenylboronic gave the desired product **1NO₂**. The aniline derivative **1NH₂** was similarly synthesised by Suzuki coupling, using 4-aminophenylboronic acid. Sonogashira coupling of **7** directly with 4-nitrophenylacetylene proved to be very low yielding. However, using TMS-acetylene as the coupling partner, followed by desilylation with TBAF, gave alkyne **8** in good yield over two steps. Subsequent coupling with 4-bromo-1-nitrobenzene gave the desired compound **2NO₂**. In contrast, 4-ethynylaniline was successfully coupled directly to the 6-bromonaphthalimide to generate the corresponding aniline analogue **2NH₂** in good yield. Acetylene **8** pleasingly also gave an intermediate to allow incorporation of the nitro-aryl functionality using CuAAC. 1-Azido-4-nitrobenzene and 4-azidoaniline were each prepared from the corresponding boronic acids *via* copper mediated conditions.^[14] CuAAC reaction of acetylene **8** with 1-azido-4-nitrobenzene and 4-azidoaniline, respectively using CuSO₄ in THF:H₂O gave the desired triazoles **3NO₂** and **3NH₂** in excellent yield for both products. Buchwald-Hartwig amination of **7** using 4-nitroaniline gave the desired product **4NO₂**. Subsequent reduction of the nitro group using Pd-catalysed transfer hydrogenation with ammonium formate as the hydrogen source, gave the aniline derivative **4NH₂** (Scheme 1).



Scheme 1: Synthesis of **1NO₂** – **4NO₂** and **1NH₂** – **4NH₂**. Reagents and conditions: (i) NBS, H₂SO₄, 60 °C, 93%; (ii) ⁿBuNH₂, EtOH, 80 °C, 55%; (iii) 4-nitrophenyl boronic acid or 4-aminophenyl boronic acid, Pd(PPh₃)₄, K₂CO₃, THF:H₂O, 70 °C, 41% (**1NO₂**), 98% (**1NH₂**); (iv) 4-ethynylaniline, Pd(PPh₃)₂Cl₂, CuI, THF:Et₃N, 80 °C, 76%; (v) 1. TMS-acetylene, Pd(PPh₃)₂Cl₂, CuI, THF:Et₃N, 80 °C, 2. TBAF, THF, 69% (over 2 steps); (vi) 4-bromo-1-nitrobenzene, Pd(PPh₃)₂Cl₂, CuI, THF:Et₃N, 80 °C, 62%; (vii) 1-azido-4-nitrobenzene or 4-azidoaniline, CuSO₄·5H₂O, NaAsc, Na₂CO₃, THF:H₂O, 66% (**3NO₂**), 98% (**3NH₂**); (viii) Pd(OAc)₂, RuPhos, Cs₂CO₃, toluene, 110 °C, 72%; (ix) Pd/C, NH₄CO₂H, EtOAc:MeOH, 98%.

Photophysical studies

The photophysical properties of **1NO₂** - **4NO₂** and **1NH₂** - **4NH₂** were next measured in CH₂Cl₂ to determine both how these changed upon reduction of the nitro group and how these varied with the nature of the linker (Figure 3, table 1). For the **NO₂** series, compounds **3NO₂**, and to a lesser extent compound **1NO₂**, show appreciable fluorescence, whereas **2NO₂** and **4NO₂** are almost completely quenched. In all four cases, reduction to the aniline resulted in a turn-on in fluorescence intensity rather than a change in emission wavelength. **4NH₂** has the weakest emission of the **NH₂** series, but also shows a significant bathochromic shift (Δ 30 nm).

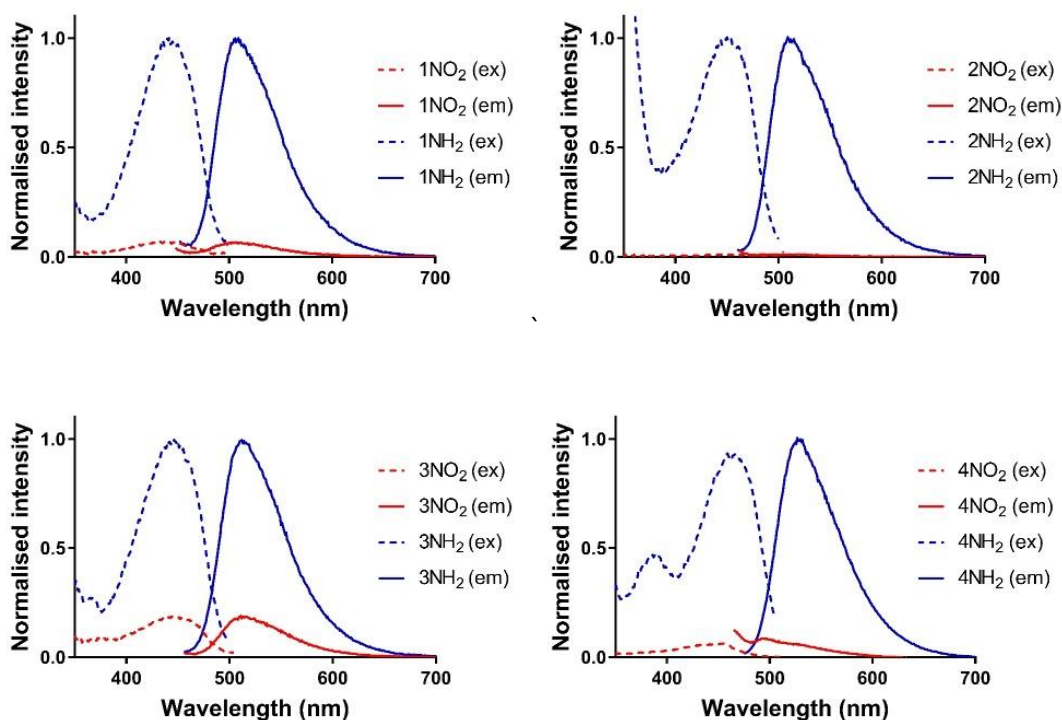


Figure 3: Excitation and emission spectra of **1** (1 μM in CH_2Cl_2 , λ_{ex} 440 nm, λ_{em} 510 nm), **2** (1 μM in CH_2Cl_2 , λ_{ex} 440 nm, λ_{em} 510 nm), **3** (1 μM in CH_2Cl_2 , λ_{ex} 440 nm, λ_{em} 510 nm), **4** (5 μM in CH_2Cl_2 , λ_{ex} 450 nm, λ_{em} 520 nm).

Table 1: Comparison of excitation and emission maxima (in CH_2Cl_2) of **1** – **4** and unsubstituted N-butyl-4-butylamino-1,8-naphthalimide.

	λ_{ex} (nm)		λ_{em} (nm)	
	NO₂	NH₂	NO₂	NH₂
Unsubstituted		421		503
1	442	440	507	509
2	445	450	520	509
3	444	446	511	512
4	456	459	497	527

Computational studies

To shed more light on the photophysical properties of the compounds, we have used first principle calculations using a technique detailed in the SI and successfully applied to many dyes before.^[15] Table 2 reports the computed positions of the lowest dipole-allowed transitions. The agreement between the theoretical and experimentally observed values is satisfactory: i) the mean absolute error is 0.20 eV, typical of such an approach;^[15a] ii) theory overshoots the experimental value, which is the expected error sign when neglecting vibronic couplings;^[15b] iii) the experimental fact that only **4NH₂** undergoes a significant bathochromic shift is reasonably reproduced.

Table 2: Comparison between theoretical (CC2, see the SI) and experimental excitation wavelengths given in eV

	1NO₂	2NO₂	3NO₂	4NO₂	1NH₂	2NH₂	3NH₂	4NH₂
Theoretical	2.94	2.89	2.93	2.93	2.99	2.95	2.94	2.69
Experimental	2.70	2.72	2.71	2.78	2.75	2.72	2.71	2.58

Figure 4 represents the density difference plots for all compounds. In all cases, except for **4NH₂**, there is a significant ICT from the *N*-butylamino group (mostly in blue, donor) at position 4 to the core of the compound (mostly in red, acceptor). The side groups added at the 6-position play a minor role. **4NH₂** is a notable exception, with a strong ICT from the amine at the 6-position to the core of the molecule, an effect well in line with the large redshift measured for that compound. In **4NO₂**, there is likely a counterbalancing effect between the nitro and amino groups, and hence, a small overall effect is evident.

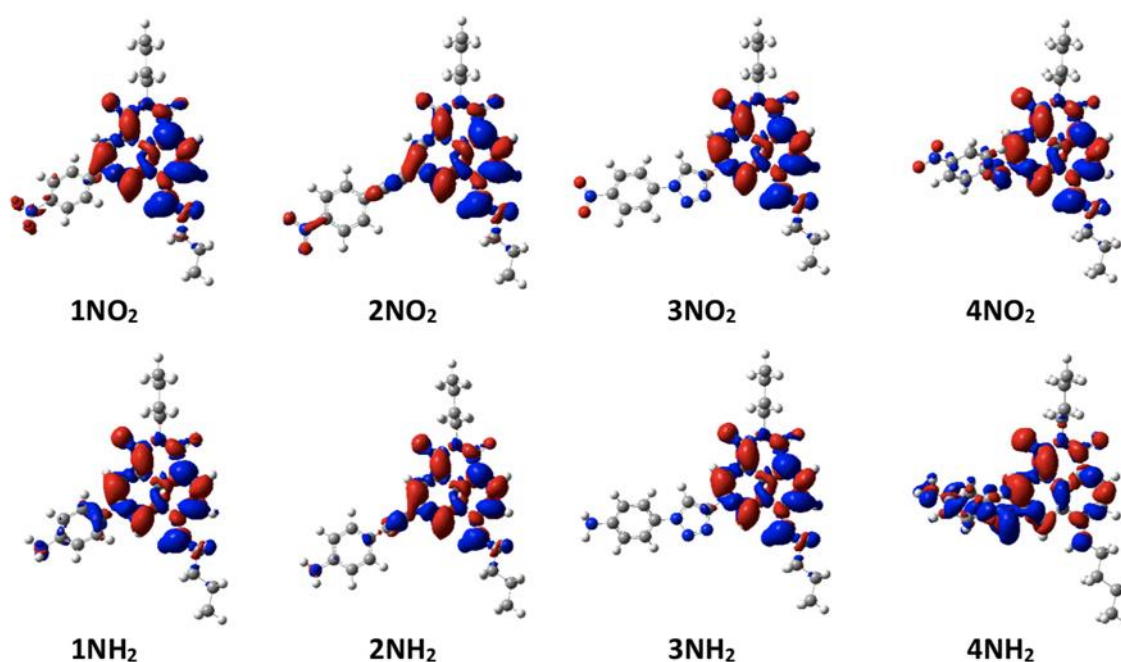


Figure 4: Density difference plots for the lowest transition in all compounds. The blue and red lobes represent regions of decrease and increase of density upon photon absorption. Contour threshold 8×10^{-4} au

Next, we examined the emission behaviour. In the **NH₂** series, the S_1 optimisations led to small structural changes in **1NH₂** – **3NH₂** with computed emission wavelengths reasonably fitting the values that were observed experimentally. Again, **4NH₂** was an exception to this general rule, as the optimisation of the lowest state led to a very low-lying “theoretical fluorescence” at 1.64 eV with a large ICT effect involving the amine at the 6-position (as in the absorption figure 4). Such a low-lying transition will be dark experimentally, due to the well-known energy gap law, and hence the very low residual emission observed experimentally is likely coming from a side effect, e.g., an emission

from S_2 or a fast emission from S_1 before its full relaxation. In that sense, the CT in **4NH₂** is too large and it consequently becomes an ineffective emitter. On the other hand, the three other **NH₂** compounds show the more classical behaviour of effective emitters with a more moderate CT, which fits with the experimental observations. As stated in the introduction, it is well known that nitro groups quench fluorescence, hence it is not surprising to see that all the **NO₂** compounds are less bright than their corresponding **NH₂** systems. Nevertheless, there are notable differences in the nitro series. For **1NO₂** – **4NO₂** the theoretical calculations predict fluorescence in the 2.52 – 2.62 eV range, which again fits the measurements (2.38 – 2.46 eV), with similar excited state topologies for all four compounds. Therefore, the experimental differences, and specifically the fact that (relative) quenching is much stronger in **1NO₂** and **2NO₂** than in **3NO₂**, should be explained by another phenomena. We reasoned that intersystem crossing (ISC) might be taking place and we have therefore computed S-T gaps on the lowest S_1 geometry for **1NO₂** – **4NO₂**. In all cases, the gap between S_1 and T_1 is very large (>0.6 eV) and therefore ISC is unlikely with the lowest triplet. In contrast, the S_1 - T_2 gaps are very low in **4NO₂** (0.00 eV) and **2NO₂** (0.07 eV) and strong quenching would be expected, as is indeed found experimentally. In **3NO₂**, T_2 lies 0.49 eV higher than S_1 making ISC very unlikely, consistent with the relatively bright fluorescence observed. **1NO₂** is intermediate with a S_1 - T_2 gap of 0.18 eV. These trends are clearly consistent with the experimental measurements.

With these results we then evaluated which scaffold would be optimal for a useful hypoxia sensor. As compound **3** had significant fluorescence in the **3NO₂** form (Φ 0.06 in EtOH, Table S1), reducing the turn on response and compound **4NH₂** was not bright enough to be a promising candidate, these scaffolds were ruled out. The turn-on response for compounds **1** and **2** made them the most promising candidates for a hypoxia probe. Non-radiative decay in 4-amino-1,8-naphthalimides is generally increased in aqueous solvents,^[16] but interestingly, the fluorescence of **2NH₂** was significantly quenched in aqueous buffer, whereas **1NH₂** remained bright (Figure 5). **1NH₂** was also the scaffold with the highest quantum yield (Φ 0.42 in EtOH, Table S1).

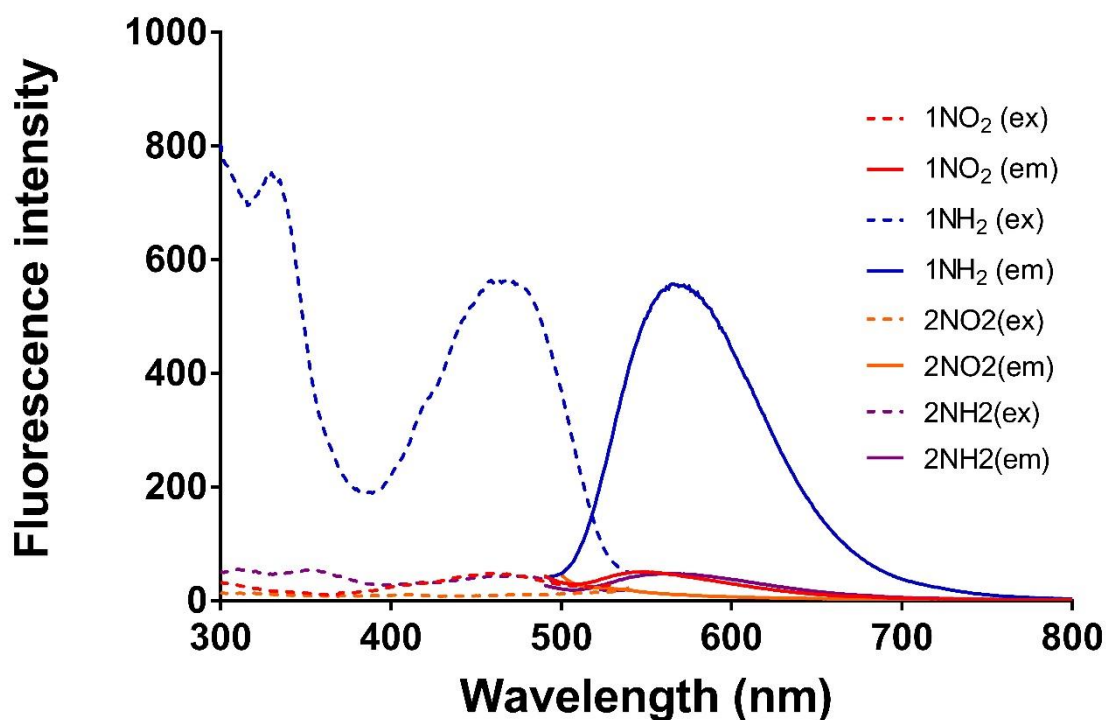


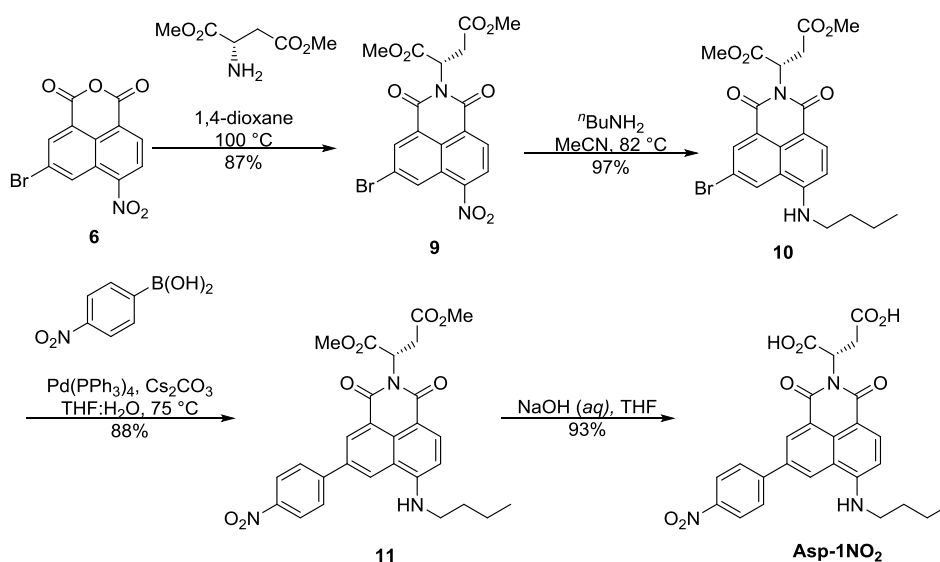
Figure 5: Excitation and Emission spectra of **1** and **2** (50 μ M in HEPES buffer 100 mM pH 7.4)

Preliminary biological studies

1NO₂ was therefore the most promising hypoxia sensor amongst those synthesised, and its potential utility was therefore evaluated in preliminary biological studies. Firstly, the cytotoxicity of **1NO₂** was assessed by Alamar blue assay, which confirmed that the compound was not toxic to DLD-1 cells (Figure S3). Next, we carried out a preliminary imaging experiment with a tumour spheroid model. DLD-1 cells were allowed to grow into three-dimensional spheroids of varying size. Such systems develop a hypoxic region which is far from the oxygenated cell culture media, mimicking the environment of a solid tumour. Imaging of spheroids incubated with **1NO₂** showed high fluorescence intensity at the periphery of the spheroid, consistent with rapid uptake of probe into the outer cells, preventing diffusion into the hypoxic region of the spheroid (Figure S4). Furthermore, the bright fluorescence intensity observed indicates that **1NO₂** is too easily reduced in the highly reducing intracellular environment, consistent with a previous work.^[8] We therefore sought to modify this system to enable imaging of the hypoxic region by developing an extracellular version of **1NO₂**, in which cellular uptake is inhibited through the incorporation of an aspartic acid group that would exist in the dicarboxylate form at biologically relevant pH.

Synthesis of extracellular analogue

We envisaged that naphthalic anhydride **6** could be functionalised selectively at the anhydride, 4-, and 6- position. This would allow for modular, orthogonal incorporation of a targeting group, sensing group, and ICT donor. We first formed naphthalimide **9** through selective condensation of anhydride **6** with aspartic acid dimethyl ester. The use of 1,4-dioxane as solvent allowed for selective reaction of the amine at the imide position, giving **9** as the sole product in good yield, with no chromatographic purification. Substitution of the nitro group upon treatment with *N*-butylamine in MeCN gave the bromide **10** in excellent yield. The bromide was then subjected to Suzuki coupling with 4-nitrophenylboronic acid to give di-ester **11**, followed by basic hydrolysis of the methyl esters to give the final extracellular hypoxia probe **Asp-1NO₂** (Scheme 2).



Scheme 2: Synthesis of extracellular hypoxia sensor **Asp-1NO₂**.

With probe **Asp-1NO₂** in hand, we first investigated the fluorescence response by chemical reduction with sodium dithionite in aqueous solution. **Asp-1NO₂** showed a strong fluorescence emission increase at 550 nm upon reduction (Figure 6). The cytotoxicity to DLD-1 cells was assessed, by Alamar blue assay, and **Asp-1NO₂** was shown to be non-toxic to cells after 24 h incubation, indicating its suitability for biological studies (Figure S3).

Asp-1NO₂ was then used in an analogous imaging experiment to that performed with **1NO₂**. Spheroids of varying size were incubated with probe **Asp-1NO₂** then imaged and a turn-on in fluorescence was observed approximately 100 μm from the spheroid surface. This distance corresponds to the limit of where oxygen can diffuse, and the beginning of the hypoxic region.^[17] In the larger spheroids, fluorescence was also observed at the very centre of the spheroid, this

corresponds to the necrotic region. In the smallest spheroid, which is lacking a hypoxic region, no fluorescence was observed demonstrating the selectivity for hypoxic microenvironments (Figure 7). The clear differences in fluorescence of **1NO₂** and **Asp-1NO₂** within spheroids (Figures S4, 7) confirm the role of the aspartic acid group in enabling the probe to travel between cells into the centre of the spheroid.

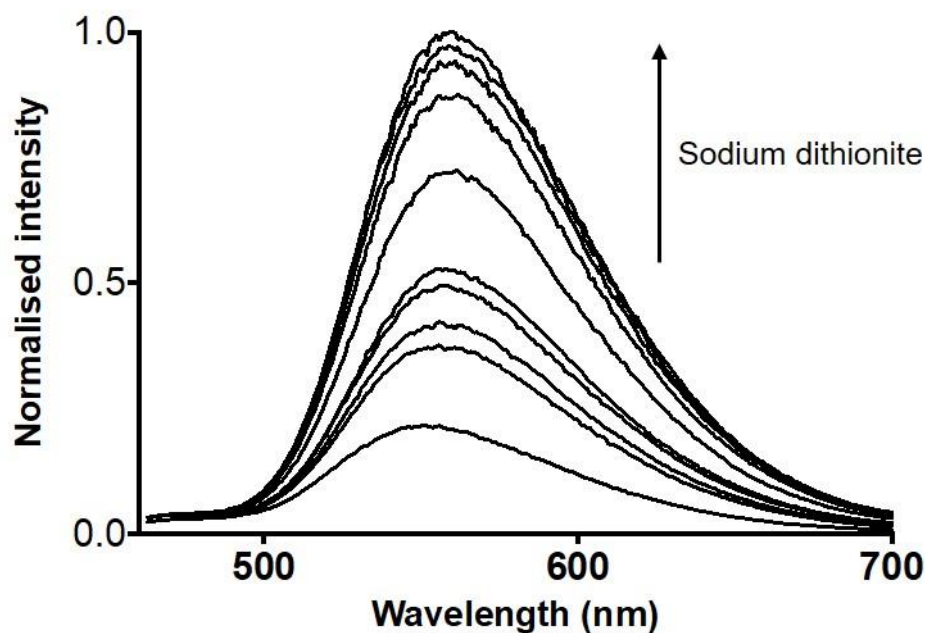


Figure 6: Fluorescence titration of **Asp-1NO₂** with increasing concentration of sodium dithionite (10 μ M in PBS pH 7.4, λ_{ex} 445 nm)

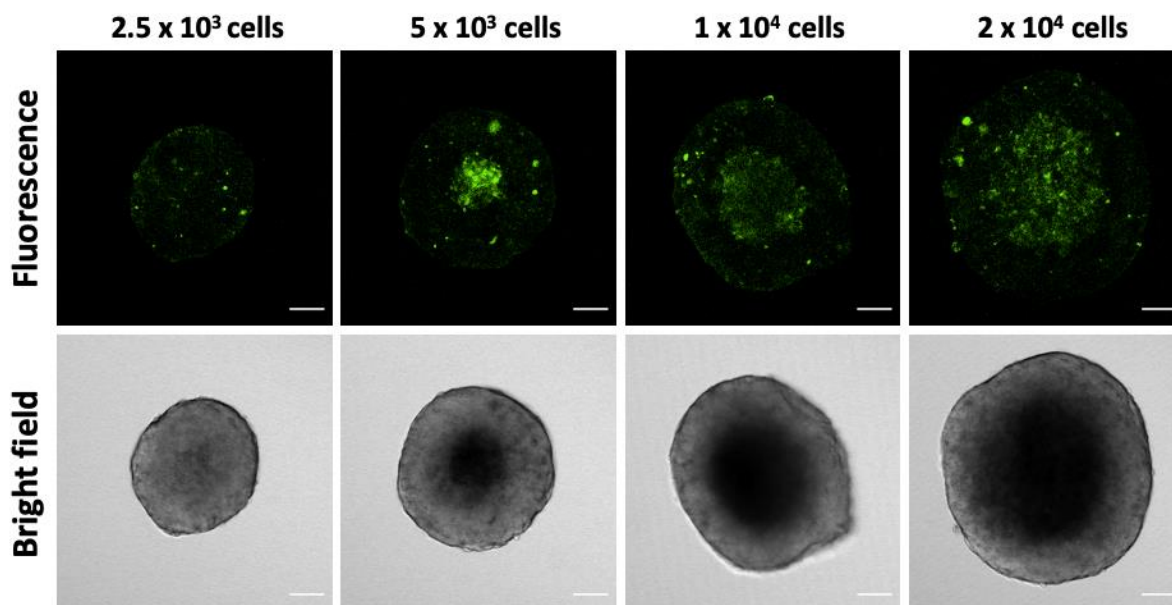


Figure 7: Confocal microscopy images of DLD-1 spheroids of varying sizes treated with 50 μ M of probe **Asp-1NO₂** (scale bar represents 100 μ m).

Conclusions:

We have demonstrated a versatile approach to the synthesis of 4-amino-1,8-naphthalimide based probes and a flexible approach to incorporation of sensing groups to the naphthalimide core using simple, modular chemistry. All linkers gave rise to a change in fluorescence intensity, and from these results we expect various sensing groups could be incorporated using an aryl-linker at the 6-position and make effective, responsive sensors. Theoretical calculations allowed us to rationalise the weak fluorescence of **4NH₂** as due to a too strong excited-state relaxation and also evidenced differences in the singlet-triplet gaps that nicely correlate with experimental fluorescence quenching for the nitro series.

For use in sensing hypoxia, the Suzuki coupled product **1NO₂** was determined to be the optimal structure, as this had the highest quantum yield, a large turn-on response upon reduction to **1NH₂**, and was the most intensely fluorescent in aqueous media. Using a modular approach, we have selectively functionalised the naphthalimide core with a targeting group at the imide position, amino donor at the 4-position, and a sensing group at the 6-position to synthesise the rationally designed extracellular hypoxia naphthalimide based probe **Asp-1NO₂**. **Asp-1NO₂** was used to successfully image the hypoxic and necrotic regions of tumour cell spheroids. One challenge for the use of green emitting fluorophores for tumour imaging, however, is their limited tissue penetration. Two-photon microscopy has been employed to address this issue for several naphthalimide based sensors, and could allow for **Asp-1NO₂** to be used for imaging in tissues.^[6c, 18] Our ongoing research also involves the development of more red-shifted probes.

This work further illustrates the versatility of the naphthalimide scaffold: we could readily substitute the group at the imide position to target any cell type or sub-cellular organelle, and easily append a variety of sensing groups. This approach to naphthalimide based probe design and synthesis is flexible allowing for the efficient generation of targeted, selective fluorescent probes and work is underway in applying this approach to the synthesis of various probes for biological purposes.

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References:

- [1] E. J. New, *ACS Sensors* **2016**, *1*, 328-333.
- [2] aS. Saha, A. Samanta, *The Journal of Physical Chemistry A* **2002**, *106*, 4763-4771; bP. Kucheryavy, G. Li, S. Vyas, C. Hadad, K. D. Glusac, *The Journal of Physical Chemistry A* **2009**, *113*, 6453-6461.
- [3] R. M. Duke, E. B. Veale, F. M. Pfeffer, P. E. Kruger, T. Gunnlaugsson, *Chemical Society Reviews* **2010**, *39*, 3936-3953.
- [4] aK. Hanaoka, Y. Muramatsu, Y. Urano, T. Terai, T. Nagano, *Chemistry – A European Journal* **2010**, *16*, 568-572; bZ. Xu, K.-H. Baek, H. N. Kim, J. Cui, X. Qian, D. R. Spring, I. Shin, J. Yoon, *Journal of the American Chemical Society* **2010**, *132*, 601-610; cR. K. Jackson, Y. Shi, X. Yao, S. C. Burdette, *Dalton Transactions* **2010**, *39*, 4155-4161; dZ. Xu, J. Yoon, D. R. Spring, *Chemical Communications* **2010**, *46*, 2563-2565; eZ. Xu, Y. Xiao, X. Qian, J. Cui, D. Cui, *Organic Letters* **2005**, *7*, 889-892; fR. Parkesh, T. Clive Lee, T. Gunnlaugsson, *Org Biomol Chem* **2007**, *5*, 310-317.
- [5] aZ. Xu, Y.-Y. Ren, X. Fan, S. Cheng, Q. Xu, L. Xu, *Tetrahedron* **2015**, *71*, 5055-5058; bS. N. Berry, V. Soto-Cerrato, E. N. W. Howe, H. J. Clarke, I. Mistry, A. Tavassoli, Y.-T. Chang, R. Pérez-Tomás, P. A. Gale, *Chemical Science* **2016**, *7*, 5069-5077; cX. Sun, S. D. Dahlhauser, E. V. Anslyn, *Journal of the American Chemical Society* **2017**, *139*, 4635-4638; dF. M. Pfeffer, A. M. Buschgens, N. W. Barnett, T. Gunnlaugsson, P. E. Kruger, *Tetrahedron Letters* **2005**, *46*, 6579-6584; eJ. F. Zhang, C. S. Lim, S. Bhuniya, B. R. Cho, J. S. Kim, *Organic Letters* **2011**, *13*, 1190-1193.
- [6] aD. Srikun, E. W. Miller, D. W. Domaille, C. J. Chang, *Journal of the American Chemical Society* **2008**, *130*, 4596-4597; bX. Sun, Q. Xu, G. Kim, S. E. Flower, J. P. Lowe, J. Yoon, J. S. Fossey, X. Qian, S. D. Bull, T. D. James, *Chemical Science* **2014**, *5*, 3368-3373; cJ. Fan, Z. Han, Y. Kang, X. Peng, *Scientific Reports* **2016**, *6*, 19562; dL. Zhang, W.-q. Meng, L. Lu, Y.-S. Xue, C. Li, F. Zou, Y. Liu, J. Zhao, *Scientific Reports* **2014**, *4*, 5870; eX.-L. Liu, X.-J. Du, C.-G. Dai, Q.-H. Song, *The Journal of Organic Chemistry* **2014**, *79*, 9481-9489.
- [7] K. G. Leslie, D. Jacquemin, E. J. New, K. A. Jolliffe, *Chem-Eur J* **2018**, *24*, 5569-5573.
- [8] K. Yang, K. G. Leslie, S. Y. Kim, B. Kalionis, W. Chrzanowski, K. A. Jolliffe, E. J. New, *Org Biomol Chem* **2018**, *16*, 619-624.
- [9] E. Collado-Fregoso, J. S. Zugazagoitia, E. F. Plaza-Medina, J. Peon, *J Phys Chem A* **2009**, *113*, 13498-13508.
- [10] R. B. P. Elmes, *Chemical Communications* **2016**, *52*, 8935-8956.
- [11] K. L. Eales, K. E. R. Hollinshead, D. A. Tennant, *Oncogenesis* **2016**, *5*, e190-e190.
- [12] aJ. Pouysségur, F. Dayan, N. M. Mazure, *Nature* **2006**, *441*, 437-443; bM. Höckel, P. Vaupel, *JNCI: Journal of the National Cancer Institute* **2001**, *93*, 266-276.
- [13] X. Sun, G. Niu, N. Chan, B. Shen, X. Chen, *Molecular Imaging and Biology* **2011**, *13*, 399-410.
- [14] K. D. Grimes, A. Gupte, C. C. Aldrich, *Synthesis-Stuttgart* **2010**, 1441-1448.
- [15] aD. Jacquemin, I. Duchemin, X. Blase, *Journal of Chemical Theory and Computation* **2015**, *11*, 5340-5359; bA. D. Laurent, C. Adamo, D. Jacquemin, *Physical Chemistry Chemical Physics* **2014**, *16*, 14334-14356.
- [16] D. Yuan, R. G. Brown, *The Journal of Physical Chemistry A* **1997**, *101*, 3461-3466.
- [17] A. J. Franko, R. M. Sutherland, *Radiation Research* **1979**, *79*, 439-453.
- [18] aB. Zhu, P. Li, W. Shu, X. Wang, C. Liu, Y. Wang, Z. Wang, Y. Wang, B. Tang, *Analytical Chemistry* **2016**, *88*, 12532-12538; bZ.-R. Dai, G.-B. Ge, L. Feng, J. Ning, L.-H. Hu, Q. Jin, D.-D. Wang, X. Lv, T.-Y. Dou, J.-N. Cui, L. Yang, *Journal of the American Chemical Society* **2015**, *137*, 14488-14495.

